IN THE SPECIFICATION:

Please amend the paragraph beginning at page 75, line 1 as follows:

The PCR reaction was carried out according to van Eijk et al. (1999), using the solid phase cDNA as template and Taq polymerase (Pharmacia Biotech, Hong Kong). Alternatively, the PCR reaction mixture contained 1x PCR buffer, each primer at 0.2μM, 0.2μM dNTPs, 1u Taq DNA Polymerase (Gibco) or 1u Tfl DNA Polymerase (Promega, Madison, WI) and 1.5 μM Mg^{+2} in a final volume of 25μl. As a control for mRNA quality, beta-actin transcripts were assayed using the same RT-PCR. PCR primers were synthesized by Besatec or Pacific Oligos (Adelaide, Australia). The following primers were used:

Gene		Primers	Product size
PAX-6	Forward: Reverse:	5'AACAGACACAGCCCTCACAAACA3' (SEQ ID NO:1) 5'CGGGAACTTGAACTGGAACTGAC3' (SEQ ID NO:2)	274 bp
Nestin	Forward: Reverse:	5'CAGCTGGCGCACCTCAAGATG3' (SEQ ID NO:3) 5'AGGGAAGTTGGGCTCAGGACTGG3' (SEQ ID NO:4)	208 bp
Oct-4	Forward: Reverse:	5'-CGTTCTCTTTGGAAAGGTGTTC (SEQ ID NO:5) 3'-ACACTCGGACCACGTCTTTC (SEQ ID NO:6)	320 bp
beta-actin	Forward: Reverse:	5'-CGCACCACTGGCATTGTCAT-3' (SEQ ID NO:7) 5'-TTCTCCTTGATGTCACGCAC-3' (SEQ ID NO:8)	200 bp
beta-actin	Forward: Reverse:	5'-TCACCACCACGGCCGAGCG-3' (SEQ ID NO:9) 5'-TCTCCTTCTGCATCCTGTCG-3' (SEQ ID NO:10)	291bp
CD-34	Forward: Reverse:	5'-TGAAGCCTAGCCTGTCACCT-3' (SEQ ID NO:11) 5'-CGCACAGCTGGAGGTCTTAT-3' (SEQ ID NO:12)	200 bp
FLK-1	Forward: Reverse:	5'-GGTATTGGCAGTTGGAGGAA-3' (SEQ ID NO:13) 5'-ACATTTGCCGCTTGGATAAC-3' (SEQ ID NO:14)	199 bp
AC-133	Forward: Reverse:	5'-CAGTCTGACCAGCGTGAAAA-3' (SEQ ID NO:15) 5'-GGCCATCCAAATCTGTCCTA-3' (SEQ ID NO:16)	200bp
Hnf-3α	Forward: Reverse:	5'-GAGTTTACAGGCTTGTGGCA-3' (SEQ ID NO:17) 5'-GAGGGCAATTCCTGAGGATT-3' (SEQ ID NO:18)	390 bp
AFP	Forward:	5'-CCATGTACATGAGCACTGTTG-3' (SEQ ID NO:19)	340 bp

	Reverse:	5'-CTCCAATAACTCCTGCTATCC-3' (SEQ ID NO:20)	
transferin	Forward: Reverse:	5'-CTGACCTCACCTGGGACAAT-3' (SEQ ID NO:21) 5'-CCATCAAGGCACAGCAACTC-3' (SEQ ID NO:22)	367 bp
Amylase	Forward: Reverse:	5'-GCTGGGCTCAGTATTCCCCAAATAC-3' (SEQ ID NO:23) 5'-GACGACAATCTCTGACCTGAGTAGC-3' (SEQ ID NO:24)	490bp
α1 anti trypsin	Forward: Reverse:	5'-AGACCCTTTGAAGTCAAGGACACCG-3' (SEQ ID NO:25) 5'-CCATTGCTGAAGACCTTAGTGATGC-3' (SEQ ID NO:26)	360bp
Keratin	Forward: Reverse:	5'-AGGAAATCATCTCAGGAGGAAGGGC-3' (SEQ ID NO:27) 5'-AAAGCACAGATCTTCGGGAGCTACC-3' (SEQ ID NO:28)	780bp

Please amend the paragraph beginning at page 79, line 21 as follows:

Differentiation into astrocyte and oligodendrocyte cells was also confirmed at the mRNA level. To induce differentiation to these lineages, spheres were plated on poly-D-lysine and fibronectin and cultured for 2 weeks in the serum free medium supplemented with EGF, bFGF and PDGF-AA. The differentiating spheres were then further cultured two weeks without growth factors and in the presence of T3. RT-PCR was used as above to demonstrate the expression of GFAP and the *plp* gene. GFAP transcripts were assayed using the following primers: 5'-TCATCGCTCAGGAGGTCCTT-3' (SEQ ID NO:29) (forward) and 5'-CTGTTGCCAGAGATGGAGGTT-3' (SEQ ID NO:30) (reverse), band size 383bp. The primers for the analysis of *plp* gene expression were 5'-CCATGCCTTCCAGTATGTCATC-3' (SEQ ID NO:31) (forward) and 5'-GTGGTCCAGGTGTTGAAGTAAATGT-3' (SEQ ID NO:32) (reverse). The *plp* gene encodes the proteolipid protein and its alternatively spliced product DM-20 which are major proteins of brain myelin. The expected band size for *plp* is 354bp and for DM-20 is 249bp (Kukekov et al., 1999). As a control for mRNA quality, beta-actin transcripts were assayed using the same primers as above. Products were analysed on a 2% agarose gel and visualised by ethidium bromide staining.